28

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1 **Review Article** 2 **Origins of Grape and Wine Aroma.** 3 Part 2. Chemical and Sensory Analysis Anthony L. Robinson,^{1,2} Paul K. Boss,³ Peter S. Solomon,⁴ Robert D. Trengove,¹ Hildegarde Heymann,⁵ Susan E. Ebeler⁵* 4 5 6 ¹Separation Science and Metabolomics Laboratory, Murdoch University, Murdoch, WA 6150, Australia, 7 ²Treasury Wine Estates, P.O. Box 396, Nuriootpa, SA 5355, Australia, ³CSIRO Plant Industry, P.O. Box 8 350 Glen Osmond, SA 5064, Australia, ⁴Plant Science Division, Research School of Biology, Australian 9 National University, Canberra, ACT 0200, Australia, and ⁵Department of Viticulture and Enology, 10 University of California, Davis, CA 95616. 11 *Corresponding author (seebeler@ucdavis.edu; tel: 530-752-0696; fax: 530-752-0382) 12 Acknowledgments: The authors thank the Australian Grape and Wine Research and Development 13 Corporation and the Australian-American Fulbright scholarship program for financial support for ALR. 14 Manuscript submitted Oct 2013, revised Dec 2013, accepted Dec 2013 15 Copyright © 2014 by the American Society for Enology and Viticulture. All rights reserved. 16 17 Abstract: Part 1 of this review summarized the current state of knowledge with respect to the 18 chemical compounds contributing to grape and wine aroma. Much of our understanding of the 19 chemistry of grape and wine composition comes from advances in analytical and sensory 20 methods for identifying and quantifying the compounds that contribute to flavor. Therefore, Part 21 2 of this review provides an overview of the chemical and sensory analysis approaches that have 22 been used to deconstruct wine flavor into its component parts with an aim toward relating the 23 chemical composition to the unique sensory properties that are associated with different wine 24 varieties and styles. 25 **Key words:** gas chromatography-mass spectrometry, multidimensional separations, sample 26 preparation, sample extraction, GC-olfactometry, sensory descriptive analysis 27

Introduction

Part 1 of this review provided an overview of the chemical components in grapes and wines and the viticultural, winemaking, and storage practices that influence their formation and concentrations. In Part 2 we provide a summary of the analytical chemistry and sensory approaches for assessing wine flavor. As noted previously, we have attempted to be as complete

33 as possible, however, given the numerous publications in these areas, not all research can be 34 covered (e.g., a Google Scholar search for the phrase "analysis of wine flavor" returns >6,600 35 publications from the years 2012–2013; the phrase "sensory analysis of wine flavor" returns 36 >5,600 publications over the same period). Therefore, we have chosen to focus on selected recent 37 applications to demonstrate the power and types of information that can be obtained with current 38 analytical and sensory approaches. The reader is also referred to several reviews for more detailed 39 discussions of selected topics (Francis and Newton 2005, Lesschaeve 2007, Plutowska and 40 Wardencki 2008, Polášková et al. 2008, Ebeler and Thorngate 2009, Muñoz-González et al. 2011, 41 Ebeler 2012).

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Chemical Analysis of Aroma and Flavor Compounds

43 Essential to understanding the various influences on wine flavor is the ability to purify. 44 identify, and quantify the compounds responsible for the sensory attributes experienced by a 45 wine consumer. Multiple techniques have been used for characterizing the composition of wine, 46 including flame atomic absorption spectrophotometry and flame atomic emission 47 spectrophotometry (Frías et al. 2003), inductively coupled plasma mass spectrometry (Baxter et 48 al. 1997), liquid chromatography (LC) (Bellomarino et al. 2009), gas chromatography (GC) 49 (Marengo et al. 2002), UV, visible, near-infrared, and mid-infrared spectroscopy (Liu et al. 2006, 50 Cozzolino et al. 2010), nuclear magnetic resonance spectroscopy (Brescia et al. 2002), and 51 electronic nose (Cynkar et al. 2010). Because GC can separate the volatile compounds in a 52 complex mixture, providing quantitative information as well as the ability to tentatively identify 53 compounds based on their retention times (which reflect the boiling point and polarity of the 54 analytes), the majority of studies assessing volatile compounds that contribute to aroma have 55 used GC methods.

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Gas Chromatography and GC-Mass Spectrometry

57 Gas-liquid chromatography (GC) was developed in 1952 (James and Martin 1952). Initial GC 58 separations used relatively short (3–10 m length, ~2 mm i.d.), packed columns that separated 59 only a few compounds in complex mixtures. For example, an early wine application separated 10 60 fusel alcohols in wine distillates (Webb and Kepner 1961). The introduction of fused-silica 61 capillary columns in 1979 (Dandeneau and Zerenner 1979) provided increased chemical

inertness and the ability to reproducibly produce very long (30–100 m) and very narrow diameter
 (0.25–0.32 mm) columns. This resulted in significantly improved efficiencies and the ability to
 reliably separate hundreds of compounds, many present at trace levels.

65 Numerous GC detectors are available, including flame ionization, which detect all molecules 66 containing reduced carbon (i.e., -CH-); nitrogen phosphorous, which specifically detect only 67 molecules containing N or P; sulfur chemiluminescence or flame photometric, which specifically 68 detect only molecules containing S; and mass spectrometer (MS), which detect ionized 69 molecules based on the ratio of their mass to charge. GC-MS has significant advantages for 70 compound identification relative to other GC detectors and analytical techniques due to the 71 availability of extensive mass spectral and retention index databases (Stein 1999, Babushok 72 2007). This, in addition to the fact that the costs of bench-top and user-friendly mass 73 spectrometers have become more affordable for the majority of users, has made GC-MS analysis 74 among the most widely used methods for analysis of wine flavors.

75 Mass spectrometry was discovered by J.J. Thomson in the early 20th century and was 76 developed by F.W. Aston in 1919, who demonstrated the existence of isotopes in non-radioactive 77 elements (Aston 1919). Modern commercial GC-MS instrumentation combines compound 78 ionization, resulting in unique mass spectral fragmentation patterns, with high-resolution 79 separation of the resulting ions, and selective and sensitive mass detection. Numerous reviews 80 discuss various aspects of wine composition with an emphasis on the role that GC-MS analysis 81 has played in contributing to current knowledge in the field of wine and grape chemistry 82 (Schreier et al. 1976, Ebeler 2001, 2012, Ebeler and Thorngate 2009, Hayasaka et al. 2005, 83 Polášková et al. 2008).

Contemporary MS detectors include various instrument configurations and types of mass analyzers, including transmission quadrupole, ion trap, and time of flight. Tandem MS (MS/MS) detectors also offer distinct advantages for sensitive and targeted analysis of known compounds.

Transmission quadrupole mass analyzers use four parallel, hyperbolic rods (electrodes) (Murray et al. 2013). Opposing pairs are connected to each other and a radio frequency and direct current voltage are applied alternately across the rods. Ions oscillate in the field with a frequency dependent on the ratio of their mass to charge (m/z). As the radio frequency voltage is varied, only ions of a specific m/z will have a stable trajectory through the quadrupoles to the

92 detector. Quadrupole ion traps (or Paul ion traps) are similar, with two ring-shaped electrodes 93 (rather than cylindrical rods) and two end-cap electrodes (Murray et al. 2013). A radio frequency 94 field is applied and ions are spatially confined within the electrodes with a cyclic motion. By 95 scanning the radio frequency field, ions of a given m/z are excited, ejected through a small 96 aperture in one end of the cap, and detected. Most guadrupole analyzers provide the ability to 97 separate ions based on a mass difference of 1 amu. With time-of-flight (TOF) mass analyzers, 98 ions leaving the GC interface and source are accelerated to the same kinetic energy. The ions 99 then travel through a flight tube with a velocity that is proportional to the m/z of the ion: that is, 100 ions with different m/z will travel at different speeds. TOF-MS instruments may provide unit 101 mass resolving powers similar to those of quadrupole instruments; however, high-resolution 102 TOF-MS instruments are also available that provide the ability to distinguish the mass of ions 103 with much greater accuracy (e.g., ethyl acetate m/z 88.1051 and 1-pentanol m/z 88.1482). This 104 can significantly aid compound identification. Fourier-transform ion cyclotron resonance (FT-105 ICR) mass spectrometers offer even greater mass accuracy. With these instruments, ions leaving the source are captured in a magnetic field where they rotate with an angular frequency that is 106 107 proportional to the magnetic field and inversely proportional to the ion mass (Murray et al. 108 2013). All ions are detected simultaneously and ions with different cyclotron frequencies (i.e., 109 different m/z) are extracted mathematically through a Fourier-transform calculation.

110 The majority of wine flavor research applications have used unit mass resolution instruments 111 (transmission quadrupoles, ion traps, or low-resolution TOF-MS). However high-resolution GC-112 TOF-MS instruments are now commercially available and the number of published applications 113 should increase over the next several years. The high cost of FT-ICR-MS has limited their 114 application for wine flavor research, although several recent articles have emphasized the 115 potential of this technique for identification of novel compounds, nontargeted profiling, and 116 authentication (Cooper and Marshall 2001, Gougeon et al. 2009, Liger-Belair et al. 2009, 117 Cuadros-Inostroza et al. 2010, Marchal et al. 2011).

Tandem mass spectrometry (MS/MS) uses more than one mass separation/analysis step, with mass separation occurring sequentially either in space or in time (e.g., ion trap instruments). In MS/MS analysis, ions from the first mass analyzer (MS1) enter into a reaction cell where they are bombarded with a reaction gas resulting in fragmentation of the ions (referred to as collision

induced dissociation, CID, or collision-activated dissociation, CAD). The second mass analyzer (MS2) is then set to analyze selected fragments resulting from the fragmentation in the reaction cell. Several types of experiments can be performed depending on whether MS1 and MS2 are set to scan a range of m/z or to filter and detect only ions of a given m/z (see review by de Hoffmann 1996). Because each mass analyzer can be set to selectively analyze only peaks of a given m/z, effectively filtering out noise and nontarget analytes, MS/MS can be a highly sensitive and selective method for targeted analysis and quantification of trace compounds.

129 GC-MS/MS analysis is increasingly used for targeted analysis of aroma compounds in wines 130 with particular emphasis on analysis of cork-taint related haloanisoles and volatile phenols 131 produced by Brettanomyces (Pizarro et al. 2011a, 2011b, Hjelmeland et al. 2012, Collins et al. 132 2012). In other selected applications, MS/MS has also been used for trace analysis of odor-active polyfunctional thiols, quantification of rotundone as a function of grape maturity and 133 134 winemaking practices, characterization of stereoisomers of wine lactone, analysis of furaneol and 135 homofuraneol, and screening of aroma-active aldehydes (Schneider et al. 2003, Luan et al. 2006, Schmarr et al. 2008a, Masson and Schneider 2009, Caputi et al. 2011, Mattivi et al. 2011). 136 137 MS/MS in combination with high-performance liquid chromatography (HPLC) has also been used to monitor the nonvolatile glycosidically bound aroma precursors in a number of studies 138 (Fedrizzi et al. 2009, Havasaka et al. 2010b, Capone et al. 2010, 2011, Kobayashi et al. 2011, 139 140 Wilkinson et al. 2011). In an interesting application, HPLC-MS/MS proved invaluable in the characterization of guaiacol conjugates in grapevine leaves and berries following exposure to 141 stable isotope-labeled guaiacol (d₃-guaiacol), simulating smoke contamination of the vine 142 143 (Hayasaka et al. 2010a). The MS/MS profile of the isotopic doublets associated with the 144 deuterated guaiacol conjugates enabled identification of seven different glycoside conjugates. In 145 addition, translocation of the guaiacol conjugates between leaves and berries was observed. 146 While not a comprehensive list of applications, these examples demonstrate the use of MS/MS 147 for a wide range of flavor characterization studies.

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Multidimensional Separations

149 While fused-silica, open-tubular capillary columns revolutionized GC separations, 150 significantly improving column efficiencies and resolution, coelution of peaks in compound

151 mixtures still frequently occurs. High-resolution accurate MS detectors and deconvolution 152 software that can distinguish closely eluting peaks based on statistical differences in their mass 153 spectra (Tikunov et al. 2005, Jiang et al. 2010, Furbo and Christensen 2012, Yang et al. 2013) 154 can aid in peak identifications. However, improvements in chromatographic resolution through 155 multidimensional separations have also received much attention. In early multidimensional 156 separations, a chromatographic peak from a first separation column was passed onto a second 157 column with a different stationary phase, via a switching or splitting device. Peaks that coelute 158 on the first column are separated in the second dimension due to differences in interactions with 159 the stationary phase on the second column. Often called "heart-cutting," this technique has been 160 frequently used in wine applications to separate chiral analytes by using a chiral column as the 161 second analytical column (Bouchilloux et al. 2000, Darriet et al. 2001, Fernandes et al. 2003, 162 Barba et al. 2010).

163 More recently, the development of comprehensive two-dimensional gas chromatography (GC×GC) (Liu and Phillips 1991) has received significant interest for analysis of highly complex 164 165 samples, including food, beverage, environmental, biological, and petrochemical (Adahchour et 166 al. 2008). In GC×GC analysis, the column effluent of the first analytical column is trapped in 167 small pulses and then quantitatively transferred to the second column. In this way, the entire set 168 of analytes eluting from the first column is separated in the second dimension. Pulse times must 169 be carefully selected to minimize peak splitting and to optimize transfer to the second column. 170 GC×GC offers enhanced separation efficiency, reliability in qualitative and quantitative analysis, 171 capability to detect low quantities, and information on the whole sample and its components, as 172 recently reviewed (Dallüge et al. 2003, Górecki et al. 2004, Ong and Marriott 2002).

173 GC×GC coupled with headspace analysis using solid-phase microextraction (HS-SPME) has 174 been used to isolate volatile compounds from the sample matrix in a range of foods and 175 beverages, including honey (Čajka et al. 2007), coffee (Ryan et al. 2004), cachaça (Cardeal et al. 176 2008), pepper (Cardeal et al. 2006), ginger (Shao et al. 2003), and grapes and wine (Perestrelo et 177 al. 2010, Rocha et al. 2007, Ryan et al. 2005, Ryona et al. 2008, 2009, 2010, Schmarr et al. 178 2010). The majority of these studies have used the method for targeted analysis where analytes 179 are selected prior to analysis, such as ethyl carbamate (Perestrelo et al. 2010), methoxypyrazines 180 (Ryan et al. 2005, Ryona et al. 2008, 2009, 201,) and aroma-active aldehydes (Schmarr 2008b).

Only a few publications have used the technique for nontargeted volatile profiling of grapes and wines (Robinson et al. 2011a, 2011b, 2011c, Rocha et al. 2007, Schmarr et al. 2010, Vestner et al. 2011, Weldegergis et al. 2011a, 2011b). In nontargeted profiling, the overall MS fingerprint, or pattern, is used to distinguish among samples and chromatographic peaks need not be identified before (or after) the analysis. The improved resolution of GC×GC separations enhances the ability to obtain detailed fingerprints of complex samples since individual peaks are more likely to correspond to unique analytes, ultimately aiding in compound identification.

188 In a recent example, GC×GC was used to analyze monoterpenes in grapes and 56 189 monoterpenes were identified in the Fernão-Pires variety, of which 20 were reported for the first 190 time in grapes (Rocha et al 2007). This highlighted the advantage that structured 191 chromatographic separation can provide for compound classification and confirmation of 192 compound identity. There continues to be new aroma compound discoveries in the grape and 193 wine research field, including (E)-1-(2,3,6-trimethylphenyl)buta-1,3-diene (TPB) (Cox et al. 194 2005) and 3,4,5,6,7,8-hexahydro-3,8-dimethyl-5-(1-methylethenyl)azulene-1(2H)-one, or (-)-195 rotundone (Wood et al. 2008). Therefore, it is anticipated that GC×GC will provide significant 196 advantages in the identification of new and novel compounds, which were previously unresolved 197 using traditional one-dimensional chromatography. Finally, GC×GC TOF-MS data are highly 198 data intensive, and improved methods for peak alignment, deconvolution, and compound 199 identification for large data sets are needed (Yang et al. 2013).

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Analysis of Glycoconjugates

Because glycosidically bound aroma compounds are nonvolatile, the most common method of glycoconjugate analysis has been to measure the aglycone and/or the sugar moiety separately following controlled acid or enzyme hydrolysis (Williams 1993, Günata et al. 1993). In these studies, GC-MS is most commonly used to analyze, identify, and quantify the aglycones, such as terpenes, benzenoids, and aliphatic residues. The released sugar moieties, such as mono- or diglucoside, arabinofuranoside, and rhamnopyranoside (Williams 1993), can be analyzed by HPLC or other chromatographic approaches.

There are a limited number of studies where the glycoconjugates are measured directly. Typically in these studies, the glycoconjugates are initially isolated and fractionated from the

210 matrix using low- (or atmospheric) pressure liquid chromatography, HPLC, counter-current 211 chromatography (CCC), and/or supercritical fluid extraction (SFE) (Strauss et al. 1987, 212 Winterhalter et al. 1990, Bonnländer et al. 1998, Palma et al. 2000). The isolated glycoconjugate 213 fraction is then analyzed directly by fast atom bombardment tandem MS (FAB-MS) (Marinos et 214 al. 1994), HPLC-MS and/or MS/MS (Hayasaka et al. 2010a), and matrix-assisted laser 215 desorption/ionization time-of-flight (MALDI-TOF) MS (Nasi et al. 2008). There are 216 opportunities to use other analytical techniques, including high-resolution TOF-MS and FT-ICR 217 MS. However, these techniques have not yet been used for the analysis of glycoconjugated 218 aroma compounds.

219 A rapid form of glycoconjugate analysis in wine is possible through application of the so-220 called glycosyl-glucose assay (Williams et al. 1995). In this approach, the glycosides are hydrolyzed and the released glucose and fructose are measured enzymatically. The assay is 221 222 rapid, accurate, and precise (Williams et al. 1995) and has been used for determination of aroma 223 potential in grapes (Zoecklein et al. 1998, Escalona et al. 1999). The determination of glycosyl-224 glucose provides a measure of the amount of conjugated compounds present but provides no 225 qualitative information about the speciation of either the sugars or the aroma compounds. In 226 addition, since grapes also contain significant amounts of glycosidically bound polyphenols (e.g., 227 malvidin-3-glucoside and rutin), the glycosyl-glucose assay will also measure sugars released by 228 hydrolysis of these compounds. Therefore, sample pretreatment with polyvinylpolypyrrolidone 229 (PVPP) may be used to remove the phenolic glycosides prior to analysis of aroma potential 230 (Zoecklein et al. 2000).

231

Sample Preservation and Preparation

232 Sample preparation is a critical step in any chemical analysis, and the information obtained 233 from chromatographic or MS analyses is highly dependent on the sample preparation method 234 chosen. Techniques that effectively isolate volatile aroma compounds or their precursors from 235 nonvolatile matrix components are typically of greatest interest for analysis of flavor compounds 236 and will be the main focus of this discussion. At any stage of sample preparation it is important to 237 preserve the compounds of interest; control of temperature, oxygen, and enzyme activity are keys 238 to reducing formation of oxidative or other chemical artifacts. As noted in Part 1 of this review, 239 many aroma compounds are present as glycoconjugates (or cysteinyl- and glutathionyl

conjugates), and preservation of conjugated compounds may be important in determining aroma potential. Therefore, cold storage prior to analysis is preferable to minimize the acid hydrolysis of glycoconjugates that can slowly occur at juice pH and ambient temperature (Skouroumounis and Sefton 2000). Further, enzyme hydrolysis by most native enzymes is inhibited in juice environments (Günata et al. 1993), but enzyme inhibitors can be added to the extract as a precaution (Razungles et al. 1993).

246 **Liquid extraction.** In many studies, volatile and glycosylated aroma precursors are isolated 247 from plant extracts, fruit juice, dealcoholized wine, and other liquid media either by selective 248 retention on Amberlite XAD resins (typically XAD-2) (Günata et al. 1985), on C₁₈ reversed-249 phase silica adsorbent (Williams et al. 1982), or by simple liquid/liquid extraction. These 250 techniques allow the isolation of aroma and aroma precursor compounds free of sugars and 251 organic acids (Günata et al. 1985). The compounds of interest are selectively eluted from the 252 resins/adsorbents or extracted with organic solvents of varied polarity (Mateo et al. 1997, Guyot-253 Declerck et al. 2000). The organic eluates are collected, dried, and concentrated for analysis. 254 These are simple and effective methods. However, there is little scope for automation, limiting 255 sample throughput, and the methods involve contact with potentially hazardous organic solvents, 256 which must be disposed of.

257 There has been recent interest in liquid microextraction techniques to minimize use of organic 258 solvents. These use very small amounts of solvents to extract analytes and include (1) single-259 drop microextraction (extraction solvent flows past a drop of liquid sample); (2) membrane 260 extraction (sample and solvent are separated by a porous polymeric membrane and solutes 261 partition into the organic phase based on their partition coefficients); and (3) dispersive liquid-262 liquid microextraction (extracting solvent is dispersed as fine droplets in the liquid sample; 263 analytes partition into the solvent which is then separated by density from the liquid sample). 264 Principles and applications of these techniques for wine analysis have been described (Kloskowski et al. 2007, Pena-Pereira et al. 2009, Ebeler 2012). In particular, dispersive liquid-265 266 liquid microextraction has been used for analysis of grape and wine flavor compounds, including 267 polyfunctional thiol aroma compounds, halophenols, geosmin, and methyl isoborneol (Campillo 268 et al. 2010, Fontana et al. 2010, Jofré et al. 2010, Pizarro et al. 2010, 2011a, Cortada et al. 2011).

The methods are rapid and sensitive; however, matrix interferences can be significant,particularly with red wines (Montes et al. 2009).

271 Static and dynamic HS extraction. The greatest advantage of static and dynamic HS 272 extraction methodologies is that they directly sample the volatile composition of the sample 273 headspace, which can then be directly related to the aroma of the sample. Static HS extraction 274 involves sampling the headspace, at equilibrium, typically using a syringe and injecting the 275 headspace gas directly into the GC inlet, often with cryofocusing to sharpen the early eluting 276 peaks. Dynamic HS extraction involves flushing the headspace of the sample vial with inert gas 277 over a defined time period. The liberated volatiles are usually captured in a cold trap or 278 adsorbent such as Tenax prior to injection onto a GC (Rosillo et al. 1999). Static HS extraction 279 has been used effectively to determine partition coefficients of analytes in aqueous ethanol 280 solutions (Conner et al. 1994, 1998, Athès et al. 2004). These methods are simple but have poor 281 reproducibility, are biased toward extraction of highly volatile and semivolatile compounds, have 282 limitations in detecting trace analytes, and as a result are often unrepresentative of the sample 283 composition (Ortega-Heras et al. 2002). Static and dynamic HS extractions have been shown to 284 be less sensitive and less selective methods for headspace analysis when compared to SPME 285 (Kataoka et al. 2000). However, in some cases, particularly for highly volatile compounds, static 286 HS methods can be highly sensitive, as when detection limits of 0.1 to 3 ug/L for low molecular 287 weight sulfur compounds in wines were obtained (e.g., H₂S and DMS) (Rauhut et al. 1998), 288 similar to those reported using other approaches, including HS-SPME-GC (Herszage and Ebeler 289 2011).

290 **HS-SPME.** Solid-phase microextraction (SPME) has been increasingly used in volatile 291 flavor analysis since its introduction in the 1990s (Arthur and Pawliszyn 1990, Arthur et al. 292 1992, Zhang and Pawliszyn 1993, Pan et al. 1995, Steffen and Pawliszyn 1996). The primary 293 advantage of this technique is that it combines analyte extraction and preconcentration in a 294 single step, without significant sample preparation. Sampling with the SPME fiber can occur 295 either from the headspace (HS-SPME) or from the liquid phase (direct immersion or DI-SPME), 296 although most applications for analysis of aroma volatiles sample from the headspace. A 297 number of grape and wine profiling studies have used HS-SPME to better understand the role of 298 various compounds in differentiating varieties, regions, and wine vintage (Marengo et al. 2002,

299 Câmara et al. 2007, Setkova et al. 2007b, Robinson et al. 2011c), and the technique has been 300 repeatedly documented as a sensitive, reproducible, automated method for preconcentration of 301 wine volatiles prior to analysis (Howard et al. 2005, Câmara et al. 2006, Setkova et al. 2007a). 302 Various parameters are routinely optimized in the development of HS-SPME techniques for the 303 analysis in wine of ethyl esters, acetates, acids, and alcohols (Siebert et al. 2005), monoterpenes 304 and norisoprenoids (Câmara et al. 2006), methoxypyrazines (Hartmann et al. 2002, Ryona et al. 2009), thiols, sulfides, and disulfides (Mestres et al. 1999a, 1999b), and furfural derivatives, 305 306 phenolic aldehydes, volatile phenols, and oak lactones (Carrillo et al. 2006). Most methods 307 described within the literature explore parameters such as fiber type, sample temperature, salt 308 concentration, agitation speed, and extraction time as part of method development and 309 optimization (Sala et al. 2000, Rocha et al. 2001, Silva Ferreira and Guedes de Pinho 2003, 310 Howard et al. 2005, Câmara et al. 2006, Carrillo et al. 2006, Setkova et al. 2007a, Robinson et 311 al. 2011b). This agrees with a recently published protocol for SPME method development 312 (Risticevic et al. 2010a). A recent review provides an extensive summary of the applications of SPME for grape and wine analysis (Jelen et al. 2012). The following sections address the 313 314 relevant parameters that should be considered for wine sample preparation for HS-SPME 315 volatile analyses.

SPME fiber type. SPME fibers are coated with a variety of single or mixed polymers that vary 316 317 in polarity, thickness, and length. The mechanisms of extraction differ: single and/or liquid 318 phases absorb analytes into the entire fiber coating and mixed and/or solid phases adsorb 319 analytes to the surface of the fiber coating. This has implications with respect to sensitivity and 320 time to reach extraction equilibrium (Risticevic et al. 2010a) and to influence the potential for 321 matrix components to compete with analytes for fiber adsorption/binding sites, as can occur with 322 carboxen-based polymers (Murray 2001). The polarity of the fiber coating also allows the user to 323 target specific compounds based on their affinity for the fiber, that is, based on the principle that 324 "like dissolves like" (Risticevic et al. 2010a). Common SPME sorptive phases or mixtures of 325 phases that are used for the analysis of volatile compounds in wine include polydimethylsiloxane 326 (PDMS), polyacrylate (PA), divinylbenzene (DVB), and carboxen (CAR) (Rocha et al. 2001, 327 Howard et al. 2005, Setkova et al. 2007a, Risticevic et al. 2010a).

328 Sample temperature. Increasing the sample temperature during extraction can increase 329 analyte partitioning into the headspace and thus increase the amount extracted improving the 330 sensitivity of the HS-SPME method (Risticevic et al. 2010a). However, desorption from the fiber 331 may also increase at higher temperatures, decreasing the overall analyte partitioning into the 332 SPME fiber phase (Fischer and Fischer 1997). In addition, it has been observed that elevated 333 extraction temperatures can modify monoterpenes (Varming et al. 2004, 2006, Perez-Cacho and 334 Rouseff 2008), esters, and acetates (Marais and Pool 1980, Ramey and Ough 1980, Pérez-Coello 335 et al. 2003) and release volatiles from glycosylated aroma precursors (Silva Ferreira and Guedes 336 de Pinho 2004). While no studies to date have thoroughly addressed the role of temperature in 337 modifying volatiles in a wine sample during SPME sampling at elevated temperatures, it has 338 been noted that different volatile profiles were observed from honey samples conditioned at 339 temperatures $>60^{\circ}$ C, compared to those kept at room temperature (Čajka et al. 2007). It is 340 possible that similar results could occur if wine or grape juice samples were also heated. This 341 aspect of HS-SPME method development should be more fully studied. However, when trying to 342 relate HS-SPME results to data obtained by sensory panels, it would seem sensible to use 343 temperatures either between 20 and 25°C, corresponding to room temperature tasting conditions, 344 or between 30 and 40°C, simulating mouth temperatures and aroma release in the mouth.

345 Salting-out. The salting-out, or Setschenow effect, describes how a nonelectrolyte, in this case an organic volatile compound, decreases in solubility following the addition of an 346 347 electrolyte such as sodium chloride to the solution (Mazo 2006). The addition of salt to wine 348 samples can increase the partitioning of aroma compounds into the headspace and increase their 349 extraction/partitioning onto the SPME fiber. Given that the Setschenow effect is related to the 350 preferential association of electrolytes with the solvent with respect to the nonelectrolyte solute, 351 it can be assumed that once the electrolyte reaches saturation the further addition of electrolyte 352 will not cause a greater effect. By extrapolating from the raw data presented in one study (Farelo 353 et al. 2004), it is observed that in a 13% ethanol solution at 30°C sodium chloride reaches 354 saturation at ~274 g/L. Dry white and red table wines are characterized by an alcohol content 355 ranging from 10 to 15% ethanol by volume, where sodium chloride is saturated at 292 and 262 356 g/L, respectively. Thus, sodium chloride concentrations between 250 and 300 g/L will generally 357 accommodate the alcohol content of wine products at or around ambient temperatures.

Interestingly, this falls within the frequently reported 100 and 350 g/L of added salt determined experimentally for optimal extractions of wine volatiles by HS-SPME (De La Calle García et al. 1998, Rocha et al. 2001, Azenha and Vasconcelos 2002, Rodríguez-Bencomo et al. 2002, Castro Mejías et al. 2003, Demyttenaere et al. 2003, Castro et al. 2004, Siebert et al. 2005, Câmara et al. 2006, Setkova et al. 2007a, Robinson et al. 2011c).

363 Sample agitation. Mechanical agitation plays an important role in accelerating mass transfer of molecules from the liquid into the headspace. Agitation increases mixing of molecules within 364 365 the liquid, creating a relatively homogenous mixture at any point in time (Zhang and Pawliszyn 1993). Agitation also increases the rotational velocity of the liquid, forcing the liquid toward the 366 367 sides of the container and thus increasing the liquid-gas interface surface area. The effects of 368 sample agitation on HS-SPME analyses have been reviewed (Zhang and Pawliszyn 1993). The rate of analyte diffusion (i.e., diffusive flux) from the liquid to the headspace is described by 369 370 Fick's first law (Equation 1):

$$J = -D \frac{\Delta \phi}{\Delta x}$$
 Eq. 1

where *J* is the diffusive flux, *D* is the diffusion coefficient, ϕ is the concentration, and *x* is the position. The diffusive flux of a compound is dependent on the concentration gradient and is directly proportional to the area of the gas-liquid interface. As agitation speeds increase, the surface area is maximized such that it is approximately equal to the ratio of the internal circumference of the vessel to the height (Equation 2):

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$$\Delta A \approx \frac{H}{0.5r}$$
 Eq. 2

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where ΔA is the change in surface area, *H* is the height of the vessel, and *r* is the internal radius of the vial. As a consequence, the greater the agitation speed, the greater the effective surface area for the transfer of volatiles across the gas-liquid interface. In addition, a higher level of agitation will result in a constant concentration of volatiles at the gas-liquid interface due to continuous effective mixing.

Extraction time. The SPME extraction time chosen will depend on the method objectives. If sampling of only volatiles in the headspace is desired, without influencing the partitioning of the volatiles from the liquid phase into the headspace, then short sampling times are used (~1 to 5 min) (Lloyd et al. 2011). Sampling under these conditions is similar to a static HS extraction (Jung and Ebeler 2003). However, in most cases, SPME extraction is optimized to maximize extraction from the liquid phase and sampling may be long enough to establish an equilibrium partitioning of the volatile analytes among the liquid phase, vapor phase, and fiber phase.

391 When choosing and optimizing extraction times, there are three scenarios that are generally 392 considered (Risticevic et al. 2010a). First, high-throughput methods require that the extraction 393 time be proportional to the separation and detection time requirements: for example, if the user 394 intends to complete a chromatographic run in 5 minutes (Setkova et al. 2007a), then the 395 extraction time is only likely to be as long or slightly longer than 5 minutes (taking into account 396 cool-down time). Short extraction times are usually pre-equilibrium conditions and are subject to 397 errors associated with slight variations in extraction times; thus, automation control is essential 398 for maximum reproducibility of the analysis (Risticevic et al. 2010a). Second, high-sensitivity 399 methods may require longer extraction times to establish equilibrium between the SPME fiber 400 and the headspace. In some instances, HS-SPME extraction times of 120 minutes have been used 401 for wine volatile analysis to maximize extraction efficiency (Câmara et al. 2006). Third, good reproducibility is paramount in quantitative and semiquantitative analysis; thus to minimize 402 403 variability in the amount of analyte extracted, equilibrium conditions should be used since these 404 reduce timing-related errors. The exception is when precise automation is available and the user 405 can demonstrate that the errors incurred due to timing are minimal (Risticevic et al. 2010a).

The optimal extraction time is dependent on the chemical and physical properties of the analytes. Optimal extraction times for a given analyte can be estimated from Graham's law of diffusion (Equation 3):

$$K_m = \frac{1}{2} \cdot m_m \cdot v_m^2 \qquad \qquad \text{Eq. 3}$$

409

410 where K_m is the kinetic energy of the analyte molecule, m_m is the mass of the molecule, and v_m is 411 the velocity of diffusion of the molecule. Assuming that the kinetic energy of any molecule is 412 constant at any given temperature and pressure, Equation 3 can be simplified (Equation 4):

$$v_m \propto \sqrt{\frac{1}{m_m}}$$
 Eq.4

413

This demonstrates that the diffusion of the molecule in the gaseous phase toward the SPME fiber is dependent on the molecular mass of the molecule. Consequently, it is expected that the analysis of higher molecular weight compounds would require a longer extraction time compared to low molecular weight compounds, which have faster diffusion rates in the headspace comparatively.

Thus, the optimum extraction time depends on the nature of the analysis and the analytes being measured. High-throughput analysis necessitates shorter extraction times but may forego the benefits of increased sensitivity for particular compounds; longer extraction times will have a greater likelihood of the samples reaching equilibrium between the liquid, vapor, and fiber phases, which will result in greater sensitivity and improved precision.

424 Desorption conditions. After extraction, the SPME fiber is inserted into the heated GC inlet 425 and the volatile analytes are desorbed and swept into the GC column by the carrier gas. 426 Desorption time and temperatures should be selected to (1) rapidly desorb the analytes within the 427 first ~30 seconds after injection; (2) quantitatively and completely transfer the analytes to the 428 column, minimizing carry-over on the fiber; and (3) minimize any thermal reactions that may 429 occur in the inlet. Artifact formation in the inlet has been observed in analysis of volatile sulfur 430 compounds using SPME sample preparation (Lestremau et al. 2004). High inlet temperatures 431 used for SPME desorption increased the oxidation of thiols; carboxen-based fibers also appeared 432 to increase the extent of oxidation during desorption in the inlet, with the carboxen possibly 433 acting as a catalyst (Lestremau et al. 2004). The formation of Maillard products following the 434 immediate thermal desorption of a liquid SPME sample from strawberry and apple fruit has been 435 documented (Verhoeven et al. 1997), and the study identified the importance of washing the 436 SPME fiber prior to introduction in the inlet when conducting liquid SPME analysis of samples 437 high in carbohydrates and/or amines.

438 Stir-bar sorptive extraction and HS sorptive extraction. Stir-bar sorptive extraction (SBSE), 439 or Twister, was developed in 1999 (Baltussen et al. 1999) and works on the same basic principal 440 as SPME where the analyte of interest partitions between the sample matrix and a polymeric 441 extraction phase. However, rather than coating the extracting polymer on a short (1 to 2 cm) 442 narrow-diameter (~ 0.56 to 0.64 mm) fused silica needle, the polymer is coated onto the outside 443 of a glass magnetic stir bar (10 to 20 mm length). With SBSE, sampling occurs by placing the 444 stir bar directly into the liquid phase for extraction, while HS-sorptive extraction (HSSE) 445 samples from the headspace/vapor phase. The major advantage of SBSE and HSSE over SPME 446 is that the stir bar is coated with 25 to 125 µL of sorptive phase (PDMS) compared with only 447 $\sim 0.5 \mu$ L of sorptive phase on a SPME fiber; the increased mass of the SBSE sorptive phase 448 allows for a substantial increase in sensitivity (Lancas et al. 2009). However, the SBSE/HSSE 449 coatings are currently limited with respect to sorptive phase types: PDMS and PDMS/ethylene glycol co-polymer are the only commercially available phases. In addition, specialized thermal 450 451 desorption and cryofocusing inlets and sampling stations are required on the GC-MS instruments. An early application of SBSE for wine analysis was for the sensitive measurement 452 453 of 2,4,6-trichloroanisole (TCA) contamination, and more recent applications for grape and wine 454 flavor analysis have recently been reviewed (Sponholz et al. 2001, Jelen et al. 2012).

455

Quantification of Volatiles

All sample preparation techniques will result in some selectivity and/or analyte losses during sample extraction/isolation (Flath 1977, Jennings and Filsoof 1977, Risticevic et al. 2010a). In addition, GC detector responses are often dependent on analyte chemical structure. While GC-MS detectors are often considered universal detectors responding to all chemical masses (within the defined mass range of the instrument), differences in analyte ionization and fragmentation efficiencies can result in different detector response ratios. For this reason, careful calibration is needed for accurate analyte quantification, depending on experimental objectives.

General reviews of external standard, internal standard, and standard addition calibration methods are widely available in most analytical chemistry textbooks (e.g., Christian 2007, Harris 2003), and the choice of calibration method will depend on the analyte, the matrix, the analysis throughput needs, and the amount of sample available. Internal standards are widely used for

467 correcting for matrix effects, sample preparation losses, and/or injection variability. Surrogate 468 internal standards are added at the beginning of the sample preparation or extraction steps to 469 monitor the entire analytical process. Using a surrogate internal standard, average losses of 470 ~50% were observed during preparation of samples for HS-SPME GC-MS profiling of volatiles 471 in Cabernet Sauvignon skins; by monitoring these losses with the internal standard, analyte 472 recoveries could be estimated for accurate quantification (Canuti et al. 2009). Internal standards 473 may also be added immediately before injection/analysis to account for injection variability or 474 variability in HS-SPME extraction. A HS-SPME approach where the internal standard is loaded 475 onto the fiber coating before sample extraction (rather than adding the internal standard 476 separately to each sample) has recently been proposed for the high throughput and reproducible 477 analysis of volatile, semivolatile, and nonvolatile analytes (Risticevic et al. 2010b).

Internal standards labeled with stable isotopes (e.g., ²H, ¹³C, and ¹⁵N) that match the analyte/s 478 of interest are widely used for accurate GC-MS quantification of aroma volatiles in foods and 479 480 beverages, including grapes and wines (Polášková et al. 2008, Ebeler 2012, Schieberle and 481 Molyneux 2012). Multiple internal standards are often used, as demonstrated in a recent study 482 where 29 stable isotope-matched internal standards were used to accurately quantify 31 different 483 wine components with HS-SPME GC-MS analysis (Siebert et al. 2005). Availability of stable 484 isotope standards is often limited, however, and require chemical synthesis if commercial standards are not available. A recent approach used a single uniformly labeled precursor (e.g., 485 $[U^{-13}C]$ - α -linolenic acid) to produce a suite of labeled products that were subsequently 486 characterized using reverse-isotope dilution procedures and then used for stable isotope dilution 487 488 analysis (SIDA) in volatile profiling studies (Gomez-Cortez et al. 2012). Such an approach may 489 be significantly more time and cost effective than synthesis of multiple labeled internal standards 490 but has not yet been applied to analysis of grape and wine volatiles.

491

GC-Olfactometry

492 GC-Olfactometry (GC-O), originally proposed in 1964 (Fuller et al. 1964), uses the human 493 nose as a detector for the compounds eluting from a chromatographic column (Acree et al. 494 1984). The method has been promoted as a useful tool in determining the sensory character of 495 some flavor compounds (Deibler et al. 1999, Kotseridis and Baumes 2000, Aznar et al. 2001,

496 Grosch 2001, Friedrich and Acree 2002, Plutowska and Wardencki 2008). In addition, 497 compounds with the most impact on the overall aroma can be defined through sequential 498 dilutions of an aroma extract—those compounds having the greatest impact on the aroma are 499 those that still can be detected sensorially by GC-O after numerous serial dilutions. Using this 500 process, a sample containing hundreds of aroma compounds can typically be reduced to a subset 501 of ~10 to 20 compounds that are most important to the overall aroma of the sample (Grosch 502 2001).

503 Although useful in characterizing aroma compounds and for initial investigations, GC-O may 504 not allow for the extrapolation of the sensory contribution of an aroma compound to the actual 505 wine sample (Barbe et al. 2008). This can be attributed to the fact that GC-O analysis essentially 506 evaluates the aroma of the compounds individually and separate from the sample matrix. GC-O 507 does not take into account the interaction effects that occur when aroma compound interact with 508 the nonvolatile matrix (Pineau et al. 2007, Robinson et al. 2009, Sáenz-Navajas et al. 2010, 509 2012) and with other volatile compounds (Atanasova et al. 2005b, Escudero et al. 2007, Pineau et al. 2009). As discussed later, these interactions may result in variations in the sensory 510 511 character of the mixture due to enhancement and suppression effects. GC-O also tends to focus 512 on potent aroma compounds, which chromatograph well at low concentrations, while abundant 513 compounds tend to overload the chromatographic column, resulting in broad peaks that elute in 514 the effluent over a long period of time and presenting only a fraction of the compound to the 515 operator to smell.

516 A recent modification of GC-O allows individual peaks or groups of peaks to be selectively 517 recombined as they elute off the column and smelled as a mixture (Johnson et al. 2012). The GC-518 recomposition olfactometry (GC-R) technique allows the creation of aroma reconstitutions 519 without the need for chemical standards or reductive aroma models. This technique provides a 520 holistic approach for understanding the aroma of complex mixtures and allows for additive, 521 masking, and synergistic effects of mixtures to be readily evaluated. When applied to wine 522 samples, this approach may make it possible to more fully characterize grape and wine varietal 523 characters, which are the result of interactions of many aroma compounds acting together to 524 produce the overall sensory perception.

525 Odor activity values (OAVs) are also widely used for selecting compounds in a mixture that 526 have the most impact on perceived aroma (Grosch 2001). In this approach, the concentration of 527 all aroma volatiles in the sample is determined and then combined with sensory threshold 528 information determined separately using the pure chemicals. The OAV is then determined as a 529 ratio of the measured analyte concentration in the sample to the analyte sensory threshold. Those 530 compounds with high OAVs would be expected to most directly impact the overall aroma of the 531 sample (Grosch 2001). This technique requires that analytes be accurately quantified and that 532 pure compounds be available to determine sensory thresholds, which, in many cases, may not be 533 possible. In addition, this approach does not account for the complex interactions that occur in 534 perception of mixtures. Since sensory thresholds are determined statistically using a large 535 number of panelists, they are dependent on a large number of variables, including the sample 536 matrix, and as such they may not reflect the range of concentrations that may actually be 537 perceived in a given food matrix by an individual. Finally, the relationship between perceived 538 intensity and concentration at suprathreshold concentrations cannot be predicted from the 539 threshold concentration (i.e., the slope of a plot of perceived intensity vs. concentration is 540 different for different compounds). Because of these limitations, OAVs are not always good 541 indicators of the contribution of individual odorants to the perceived intensity of a mixture 542 (Audouin et al. 2011), as further demonstrated with the white wine variety Maccabeo using 543 reconstitution studies with a dearomatized white wine matrix and pure aroma compounds 544 (Escudero et al. 2004). Selected compounds such as fusel alcohols, acids, and esters, even when 545 present at concentrations above their sensory thresholds, did not have a significant impact on the 546 overall wine aroma. On the other hand, a group of compounds with odor activities less than one 547 were important to reconstructing the aroma of the original Maccabeo wine.

The analytical approaches described in the above sections are focused on fully characterizing the chemical composition of grapes and wines. GC-O approaches attempt to link composition directly to the sensory impact of individual compounds. However, as discussed above with GCrecomposition olfactometry analysis, holistic measurements of sensory perception are also necessary to fully characterize wine flavor. Such approaches for sensory evaluation of wine are discussed in the following section.

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Sensory Evaluation of Wine

556 Sensory evaluation stems from the experimental psychology field of psychophysics, which 557 explores how human responses are elicited by chemical and physical stimuli. Early work in this 558 field by researchers including Ernst Weber and Gustav Fechner investigated the mathematical 559 relationship between the physical and perceptual magnitude of stimuli and is still the subject of 560 much cognitive research (Dehaene 2003). Where psychophysics might focus experimental 561 research on understanding how humans respond to stimuli, sensory evaluation focuses on using 562 human subjects to explore the sensory properties of stimuli. Sensory evaluation has been defined 563 as a scientific method used to evoke, measure, analyze, and interpret those responses to products 564 as perceived through the senses of sight, smell, touch, taste, and hearing (Lawless and Heymann 565 2010). Sensory evaluation in the food, beverage, and fragrance industries was historically 566 conducted by product experts such as the cheesemaker, winemaker, or perfumer, who had 567 training and extensive knowledge of how raw materials and processing affected the finished 568 product quality (Sidel and Stone 1993). Although these product experts are still commonly used 569 in industry today, the use of specialized sensory panels for conducting discriminative, 570 descriptive, and affective evaluations of products has many more advantages. It is generally 571 recognized that the judgments of a panel are generally more reliable than the judgments of an 572 individual, there is less potential risk that the single expert might be ill/retire/die or otherwise be 573 unavailable to make decisions, and, most importantly, the opinions of the expert may or may not 574 reflect what consumers want in a product (Sidel and Stone 1993).

575 Several types of sensory tests are possible, including discrimination, descriptive, and 576 consumer. Discrimination tests determine whether two products are perceptibly different from 577 one another, for example, due to a modification in the production process or the identification of 578 a defect. These tests are commonly fast and easy to conduct and require little training. However, 579 they provide little additional detail about why the differences exist or the relative impact of the 580 difference (Peryam and Swartz 1950, Lawless and Heymann 2010). Descriptive tests are used to 581 obtain a more detailed description of the sensory attributes of a product. These tests assist in 582 identifying which attributes vary due to a modification to the product or to compare attributes 583 between/among products. Descriptive tests typically require additional time and panel training 584 compared to difference tests (Murray et al. 2001, Lawless and Heymann 2010). Consumer

(hedonic) tests are used to determine if a consumer likes a product, prefers it to another product, or finds the product acceptable based on its sensory characteristics. These tests are different from market research where the extrinsic factors that can affect wine choice are also considered, such as brand, region, price, and awards (Lockshin et al. 2006, Lawless and Heymann 2010).

589 Wine is a highly diversified food product made from numerous grape varieties, grown in a 590 diverse range of environments worldwide, and produced using a multitude of permutations in 591 viticultural management and winemaking techniques that have been developed over the course of 592 centuries. As such, the sensory characteristics of wines are highly varied. Use of standardized 593 terminology can aid in the communication of sensory attributes of wine products among 594 winemakers, marketers, consumers, and researchers (Noble et al. 1984, 1987, Gawel 1998, 595 Gawel et al. 2000). However, standardized terminology is not always used by winemakers, wine 596 writers, and consumers, who frequently use everyday language to infer relationships between the 597 wine product and the sensory properties of other common food products and smells. This was 598 demonstrated in a study of vegetal aromas in Cabernet Sauvignon wines using expert 599 winemakers and trained sensory panelists (Preston et al. 2008). Experts used a variety of terms to 600 group and describe vegetal characters in the wines, and in some cases vegetal and sulfur 601 attributes were often grouped similarly. However, trained panelists reliably distinguished 602 between related vegetal terms such as a fresh, bell pepper-like aroma, a cooked vegetal aroma (a 603 mixture of canned potatoes corn, green been, and asparagus), a green/black olive aroma, and a 604 eucalyptus aroma. Each of these aroma attributes would arise from different chemical 605 components. Therefore, in order to minimize or optimize specific aroma characteristics in a 606 wine, precise and well-defined terminology for sensory attributes, as is obtained with sensory 607 descriptive analyses, is needed when communicating about wine sensory properties.

Descriptive analysis of wine and linking sensory and chemical data. To quote a succinct summary of the importance of descriptive sensory analyses, "Without sensory evaluation, even precise information about the volatile composition in the nasal passages cannot predict the flavor of the system as perceived by humans" (Noble and Ebeler 2002). Descriptive sensory analysis is one of the most powerful tools for conducting product comparisons and for determining relationships between sensory properties of foods and beverages and their composition or consumer liking (Murray et al. 2001, Lawless and Heymann 2010). There are a number of

615 different methods for conducting descriptive analysis, including the flavor profile method, 616 texture profile method, Quantitative Descriptive Analysis[™], the Spectrum[™] method, 617 quantitative flavor profiling, and free-choice profiling, and their comparable advantages and 618 disadvantages have been previously discussed in a review of the field (Murray et al. 2001). 619 Descriptive sensory analysis has been used extensively in the wine industry over the last 30 years 620 following pioneering work conducted by Ann Noble and coworkers at the University of 621 California, Davis, in the late 1970s and early 1980s (Arnold and Noble 1979, Schmidt and Noble 622 1983, Aiken and Noble 1984, Heymann and Noble 1987, Noble and Shannon 1987).

623 Numerous studies have used descriptive sensory analysis to explore differences in the sensory 624 characteristics of single variety wines, including Cabernet Sauvignon (Heymann and Noble 625 1987), Chardonnay (Arrhenius et al. 1996), Pinot noir (Guinard and Cliff 1987), and Zinfandel 626 (Noble and Shannon 1987) from California, Seyval blanc from Missouri (Andrews et al. 1990), 627 Cabernet franc from the Niagara region (Hakimi Rezaei and Reynolds 2010), Cabernet 628 Sauvignon from Australia (Robinson et al. 2011a), Riesling from Germany (Fischer et al. 1999), Malbec from Argentina (Goldner and Zamora 2007), Albariño from Spain (Vilanova and 629 630 Vilariño 2006), Touriga Nacional and Tinta Roriz from Portugal (Falqué et al. 2004), and Sauvignon blanc from New Zealand and other countries (Lund et al. 2009). In addition, 631 632 numerous studies have predominantly used descriptive sensory analysis to explore the sensory 633 impacts of various viticultural and enological treatments (Lesschaeve 2007, Robinson et al. 634 2011b), such as the influence of oak (Francis et al. 1992, Reynolds et al. 2001, Cano-López et al. 635 2008), fermentation with different yeast strains and temperatures (Eglinton et al. 2000, Soden et 636 al. 2000), wine storage temperature conditions (Francis et al. 1994, De La Presa-Owens and 637 Noble 1997, Robinson et al. 2010, Hopfer et al. 2012b, 2013), closure types (Godden et al. 2001, 638 Skouroumounis et al. 2005a, 2005b, Hopfer et al. 2012b, 2013), grapevine diseases including 639 Botrytis cinerea and Uncinula necator (powdery mildew) (Stummer et al. 2003, 2005, Sivertsen 640 et al. 2005), and grapevine water status, crop yield, and canopy management (Reynolds et al. 641 1996, 2013, Chapman et al. 2004, 2005).

Although many studies have explored the sensory differences among imposed treatments,
 there is much current interest in exploring the relationships between these sensory differences
 and wine composition and/or the sensory aspects of wine associated with consumer preferences

645 (Francis and Newton 2005, Lesschaeve 2007). Recent examples include identification of sensory 646 attributes that drive consumer and expert acceptance of Shiraz and Cabernet Sauvignon wines 647 (Lattey et al. 2010), assessment of the relationship between sensory and chemical data for oak-648 derived compounds found in French and Spanish wines (Prida and Chatonnet 2010), modeling of 649 wine mouthfeel attributes using metabolomic data (Skogerson et al. 2009), comparisons between 650 vine vigor status with tannin and sensory data (Cortell et al. 2008), assessment of the 651 relationships between red wine textural characteristics and the chemical composition of Shiraz 652 wines (Gawel et al. 2007), comparisons of volatile components of sweet Fiano wines and sensory 653 data (Genovese et al. 2007), and comparisons between volatile compositional data and sensory 654 data of Chardonnay, Cabernet Sauvignon, Malbec, and Sauvignon blanc wines made in different 655 styles and/or from different regions (Lee and Noble 2006, Robinson et al. 2011a, 2011b, Benkwitz et al. 2012, Hjelmeland et al. 2013, King et al. 2014). Future work may also consider 656 657 aspects of wine and food interactions, given that an understanding of how people enjoy pairing 658 wine with food combinations is commonly discussed in the popular literature (Madrigal-Galan and Heymann 2006). 659

660 In a recent study both sensory descriptive analysis and chemical analysis were used to 661 understand the influences of blending on chemical and sensory profiles of Cabernet Sauvignon, 662 Merlot, and Cabernet franc monovarietal wines and their blends (Hopfer et al. 2012a). Blending resulted in 663 changes to both sensory and chemical characteristics relative to the original monovarietal wines. The measured 664 chemical composition reflected the average composition of each mono-varietal wine in the blend. However, the 665 sensory profiles exhibited examples of suppressing or amplifying effects, and the overall sensory profile was not a 666 simple average of the sensory profile of the monovarietal base wines. Interestingly, similar sensory profiles could be 667 obtained with very different blending mixtures. This study demonstrates the value of using both sensory and 668 chemical analyses to describe wine aroma and flavor, but also points to the difficulties in fully linking compositional 669 information to sensory properties due to complex aroma interactions. These aroma interactions are further discussed 670 below.

In all cases, descriptive sensory analysis produces multivariate data in relation to a single sample set. In the above examples, multivariate statistical techniques have also been critical in exploring relationships between descriptive sensory data and compositional and/or consumer sensory data, such as multivariate analysis of variance (MANOVA), principal component analysis (PCA), canonical variate analysis (CVA), generalized Procrustes analysis (GPA), and

partial least squares (PLS) regression. In general, the use of multivariate statistics for relating
sensory and chemical data has been well described (Noble and Ebeler 2002, Lawless and
Heymann 2010, Naes et al. 2011).

679 There is still need for improved methods for handling the large data sets involved in 680 correlating analytical and sensory data. Standard workflows, particularly for analysis of 681 chromatographic data, are time-consuming and involve many steps where automated data 682 processing is still limited, as discussed previously. In addition, many of the statistical approaches 683 currently used have limitations when the number of samples or treatments is small with respect 684 to the number of variables (as frequently happens with hyphenated GC data such as GC×GC and 685 high-resolution MS where hundreds of peaks may be present in a sample). Further, relationships 686 among analyte concentrations, sensory responses, and treatment variables may not be linear, 687 making it difficult to interpret results from statistical approaches based on linear relationships. 688 Finally, in cases where predictive models are developed (e.g., PLS analysis relating analyte 689 concentrations to sensory attribute intensities), methods of cross-validation and model testing 690 must be clearly defined and the variability in responses observed within the training sets must 691 represent the variability in the samples tested. An excellent overview of these issues has recently 692 been presented (Smilde et al. 2013). An understanding of these limitations and new approaches 693 for analysis of complex multivariate data sets may provide new opportunities for linking chemical and sensory data to fully understand the flavor of complex chemical mixtures. 694

695 **Interaction effects on wine flavor.** As previously discussed, much wine sensory research has 696 focused on correlating descriptive sensory and quantitative analytical data in order to 697 successfully identify odor compounds that contribute to the overall aroma perception of wine 698 (Guth 1997a, 1997b, 1998, López et al. 1999, 2003, Kotseridis and Baumes 2000, Ferreira et al. 699 2001, 2002, Lee and Noble 2003, Escudero et al. 2004, Campo et al. 2005, Bailly et al. 2006, 700 Benkwitz et al. 2012). The use of sensory evaluation to elucidate the impact of complex aroma 701 compound interactions, including masking and enhancing effects, is likely to improve our 702 understanding of the perceived aroma of wine (Atanasova et al. 2005a). For example, using 703 descriptive analysis, it has been shown that fruity aromas in wines can significantly mask vegetal 704 characters in the wines (Hein et al. 2009). Similarly, at low concentrations, ß-damascenone has 705 the ability to mask the herbaceous aroma associated with 3-isobutyl-2-methoxypyrazine (Pineau

et al. 2007) while enhancing the berry fruit aromas in red wines (Escudero et al. 2007, Pineau et al. 2007). As noted in the above section, the aromas of wine blends exhibited suppressive and amplifying effects when the intensity of specific aroma attributes of the blends was compared to the original aroma of the monovarietal wines (Hopfer et al. 2012a). These results indicate that aroma interactions in the complex wine blend mixtures can impact the overall aroma in a nonadditive manner (Hopfer et al. 2012a).

712 The matrix components can also impact aroma volatility, release, and perception. Recent 713 sensory research has shown that ethanol suppresses the fruity notes in model wine solutions 714 (Grosch 2001, Escudero et al. 2007, Le Berre et al. 2007, King et al. 2013) due to the increased 715 solubility of the volatiles when ethanol is present (Le Berre et al. 2007, Robinson et al. 2009) and 716 to the inhibition of volatile compound odor activity by ethanol (Grosch 2001). For example, ß-717 damascenone is recognized universally as a potent wine aroma compound (Skouroumounis and 718 Sefton 2002, Pineau et al. 2007, Sefton et al. 2011); however, its reported aroma threshold varies 719 from 2 to 13 ng/L in water (Buttery et al. 1990, Czerny et al. 2008), to 50 ng/L (Guth 1997a) in 10% aqueous ethanol, to 7000 ng/L in a red wine matrix (Pineau et al. 2007). 720

721 These results indicate that in addition to ethanol, other wine components also impact aroma 722 perception, therefore understanding the factors that influence the release of volatiles from the 723 complete wine matrix is of major importance to understanding wine aroma perception (Plug and 724 Haring 1994). Several studies have shown that polyphenols, tannins, polysaccharides, and 725 proteins can interact with aroma compounds and affect their volatility and release from solution (Dufour and Bayonove 1999a, 1999b, Voilley and Lubbers 1999, Jung et al. 2000, 2002, Jung and 726 727 Ebeler 2003, Aronson and Ebeler 2004, Jones et al. 2008, Saenz-Navajas et al. 2010, 2012, 728 Villamor and Ross 2013). A recent study clearly demonstrated that nonvolatile tastants (e.g., 729 polyphenols, tannins/high molecular weight components, glycerol, and organic acids) can 730 significantly impact the perception of wine aroma compounds, enhancing the intensity of some 731 attributes and decreasing the perceived intensity of others (Frank et al. 2011). The mechanisms for 732 the interactions are not entirely clear but may be due to changes in solubility or noncovalent 733 interactions/associations of the odorants with the solute components in solution (Connor et al. 734 1998, Jung et al. 2000, 2002).

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The existence of interaction effects clearly supports the use of holistic approaches, such as descriptive analysis, in the sensory assessment of wine products where physical, chemical, and perceptual enhancement and suppression effects complicate the assessment of volatile components in isolation.

739

Summary

740 Advances in analytical, sensory, and statistical analysis have been critical for understanding 741 the relationships between grape and wine composition and sensory perception. However, it is 742 still not possible to fully predict aroma quality based on chemical composition alone, due, in 743 part, to the potential presence of trace compounds that may be difficult to quantify and identify 744 and to the complex interactions of aroma compounds with each other and with the wine matrix 745 that impact aroma volatility, release, and perception. As outlined in this review, future 746 discoveries will continue to be driven by development of improved and high-throughput 747 analytical methods that will allow monitoring of a large number of volatiles, including those 748 present at low concentrations. Sensory and statistical approaches that allow for handling of large 749 multivariate data sets are also needed. Ultimately, multidisciplinary studies using genomic, 750 proteomic, metabolomics, and sensory techniques to understand flavor and aroma formation in 751 the grape and during fermentation will generate essential knowledge about the role that 752 production processes—from the vineyard to the cellar—have on the final product presented to a 753 consumer.

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1441